

# Workflows for RNA-Seq analysis

**Best practices on managing, analyzing and visualizing RNA-Seq data**  
**A perspective from the CCR Bioinformatics Core**

**Parthav Jailwala**  
**CCR Bioinformatics Core**

# What is the CCR Informatics core ?

- Collaborative resource providing bioinformatics analysis assistance to CCR investigators
- 138 requests concluded across 91 labs
  - ✓ 29 requests concluded for RNA-Seq data analysis
- Workflow/pipeline development
- Education and training

# Value of working with CCRIFX

- Understand research question & suggest viable experimental designs
- Guide on use of technologies
- Assist in data tracking and transfer of large datasets
- Perform analysis and explain results
- Deliver figures and methods for manuscripts, presentations, posters
- Maximize research impact of your invested effort

# Outline

- Computational aspects of RNA-Seq data analysis
  - Best practices
- Example CCRIFX pipeline for RNA-Seq analysis:
  - Pre-alignment QC
  - Tuxedo RNA-Seq pipeline
  - Post alignment QC
- Visuals of FastQC plots (good vs bad data)
- Pointers

# RNA-Seq : A 5-step process

**Design Experiment**



**Purify RNA**



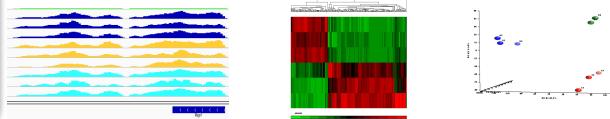
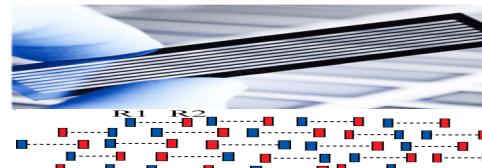
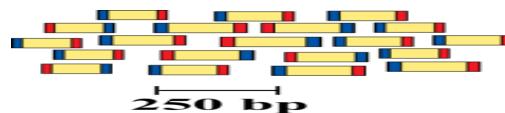
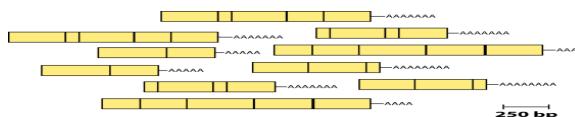
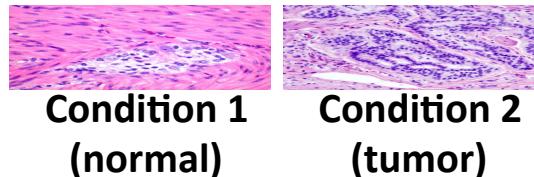
**Prepare Libraries**



**Sequence**



**Analysis**



**Primary Objective**  
Replication & Sequencing depth  
Samples of interest

Isolate RNAs  
Target enrichment  
RNA fragmentation

Generate cDNA, fragment,  
size select, add primers,  
adapters

Sequence ends

Image analysis, Base calling,  
QC, Mapping, DGE analysis,  
visualization and Biological  
interpretation

# Features of RNA-Seq data

- **Very large volume of raw and processed data**
- **Variety of file formats with non-standard naming conventions**
- **Analysis requires specialized software tools**

# Computational aspects of RNA-Seq analysis

- **Data Management Plan**
  - Data retrieval and short-term storage
  - Data backup and archival storage
  - Standardized nomenclature (file and directory naming)
- **Quality control of sequence reads & alignments**
  - Source: Low-quality RNA, library construction biases, sequencing chemistry
  - Solution: Trimming and/or filtering reads before alignment
- **Primary data analysis**
  - Pre-alignment QC
  - Alignment to the transcriptome
  - Post-alignment QC
  - Quantifying transcript levels & Differential expression detection
- **Data visualization and Biological interpretation**

# Why automate an RNA-Seq workflow ?

- Standardizes and codifies the procedure
- Increases consistency and robustness of the analysis across multiple samples or experiments
- Facilitates evaluation of changes and simplifies debugging of anomalies and artifacts
- Minimizes human error

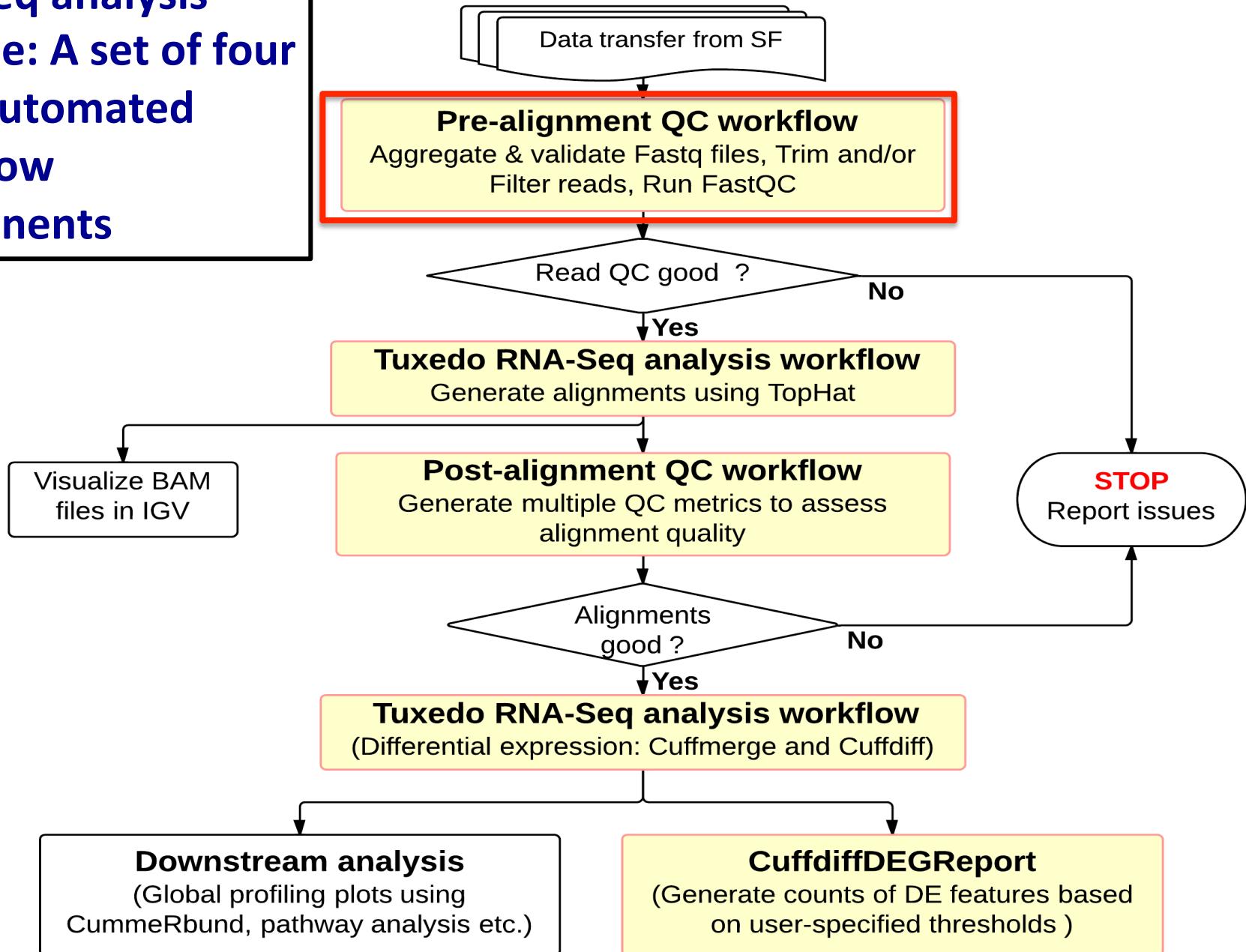
# A large RNA-Seq project that makes use of workflows

- Transcriptomes in mouse embryos & embryonic stem cells
- 39 samples: 13 conditions x 3 biological reps/condition
- HiSeq2000, 101bp PE reads, Avg. 161M reads/sample
- 10 differential expression comparisons
- 4 TB of raw fastq files, 11 TB of analysis files

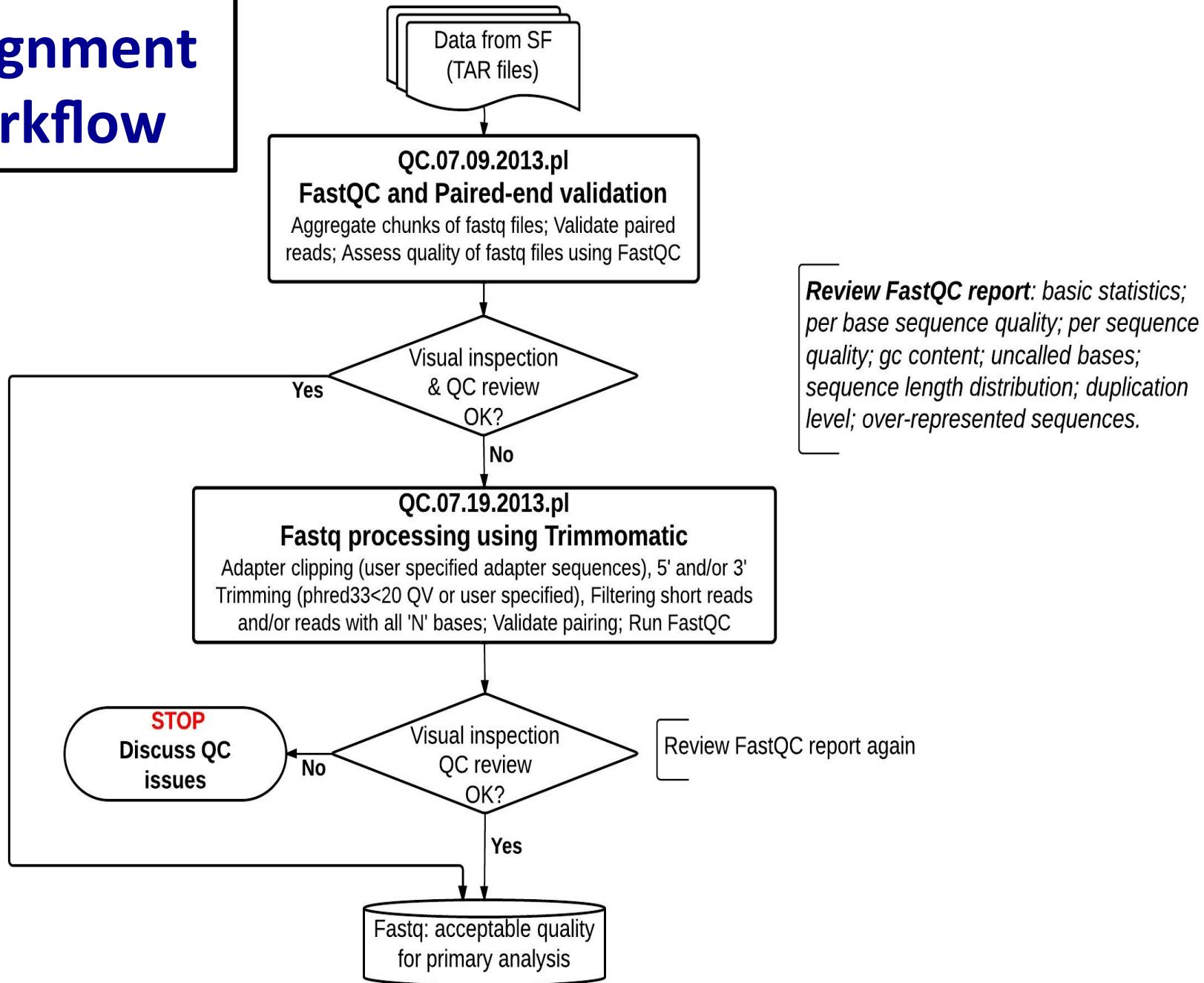
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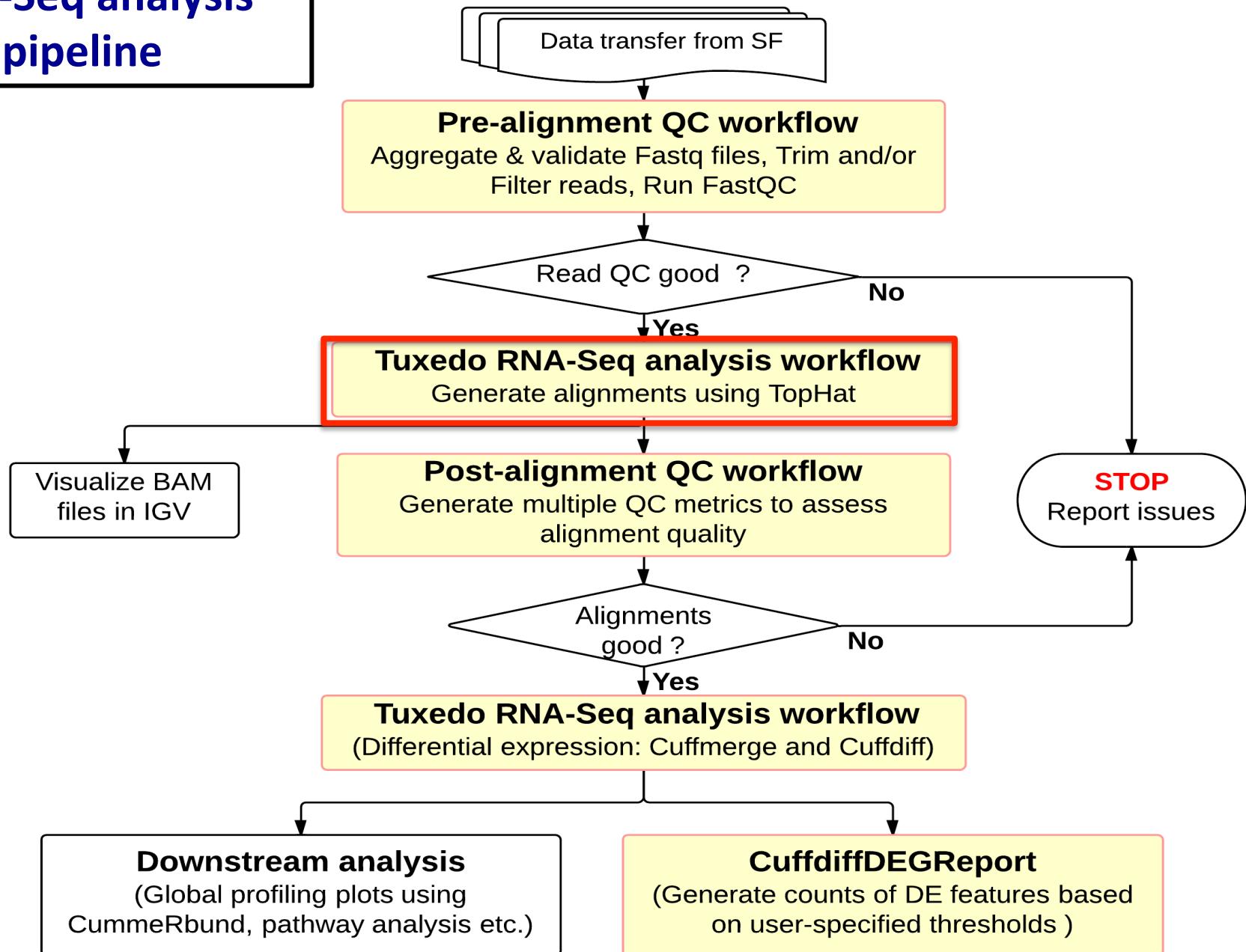
# RNA-Seq analysis pipeline: A set of four semi-automated workflow components



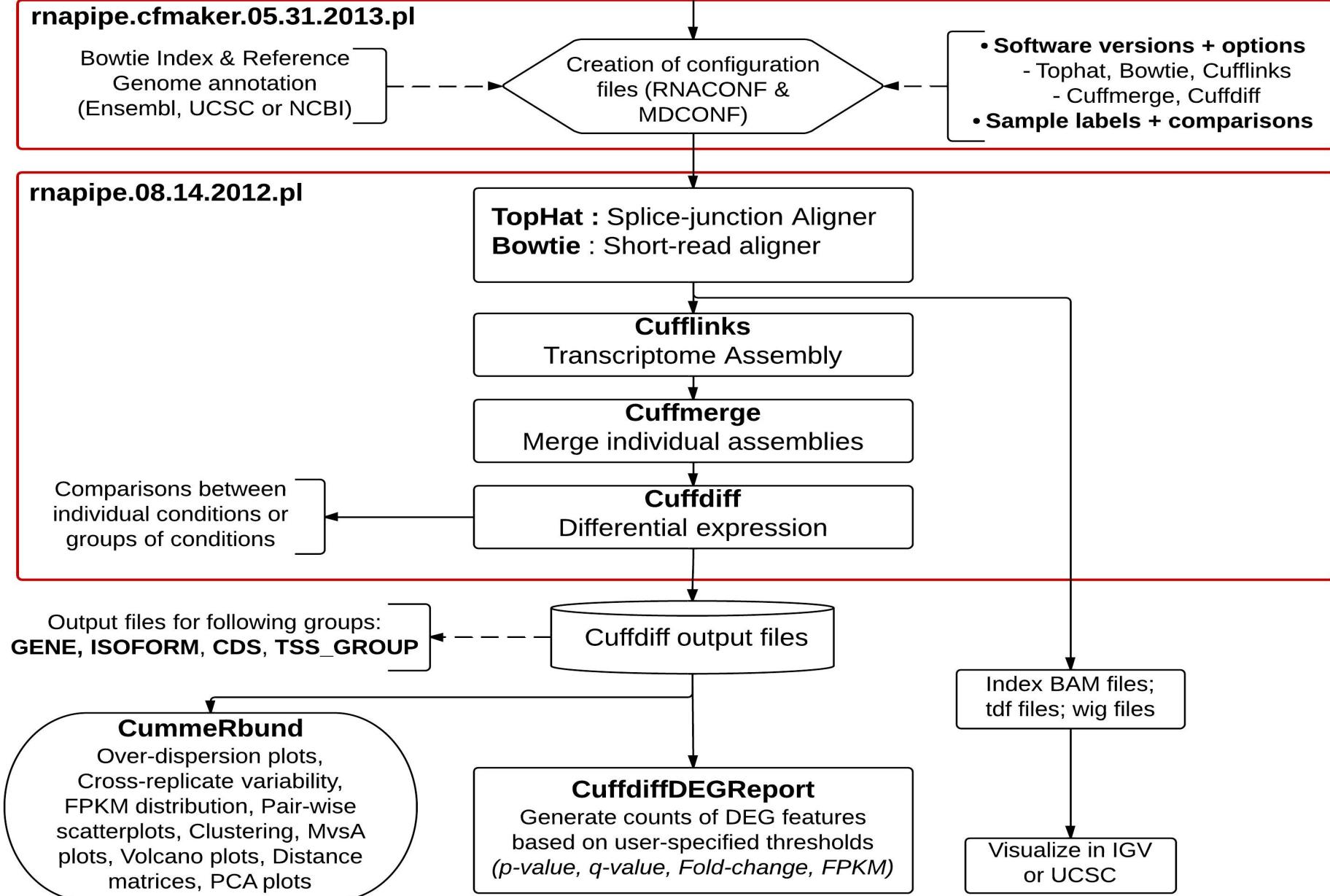
# Pre-alignment QC workflow



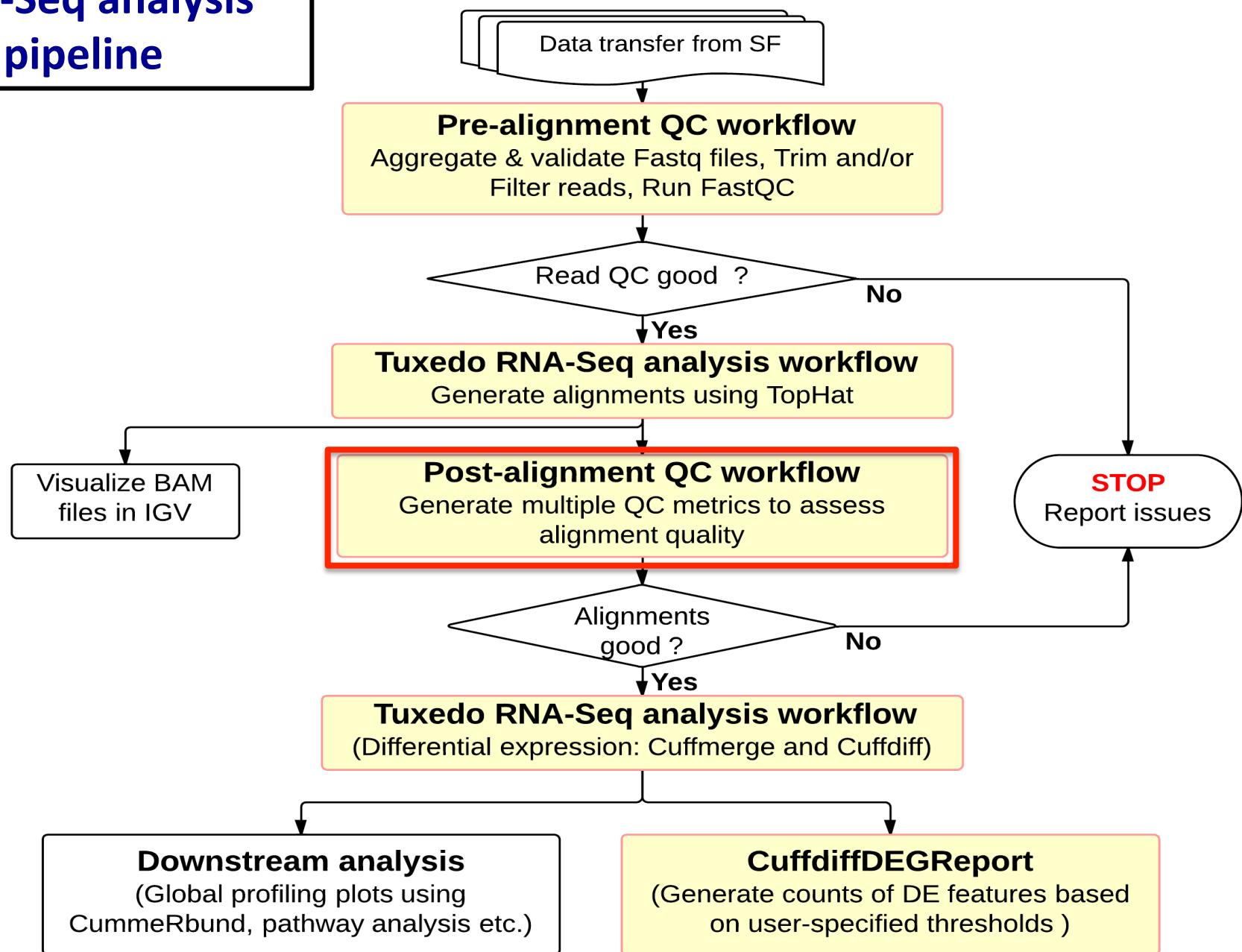
# RNA-Seq analysis pipeline



# Tuxedo RNA-Seq analysis workflow



# RNA-Seq analysis pipeline



# Post-alignment RNA-Seq QC

RNASeqQC.09.14.2012.pl

BAM files processed using Tuxedo  
RNA-Seq analysis workflow

## Pre-processing of BAM files

CleanSam, AddReplaceReadGroups, Make Sequence  
dictionary, ReOrder BAM, Sort BAM, Index BAM

Prepped.BAM

## RNA-SeQC metrics

Read count metrics, rRNA contamination, strand  
specificity, coverage metrics, GC stratification

## Picard metrics

Alignment metrics, Insert size metrics,  
Distribution across genomic features

Per sample metrics in text files  
(metrics.tsv) + HTML report

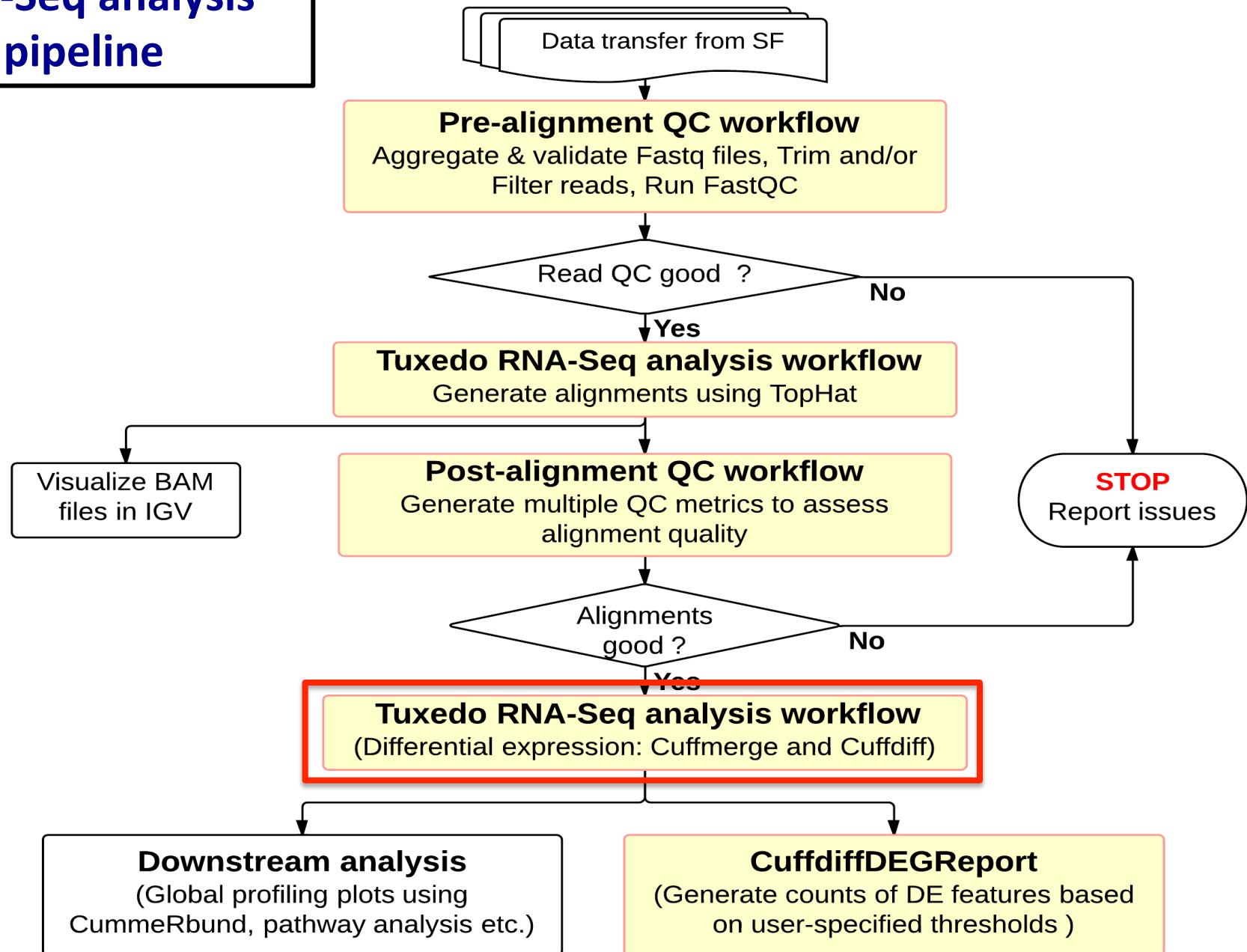
Per sample metrics in text files +  
Graphical output in PDF format

RNASeqQCReport.09.06.2012.pl

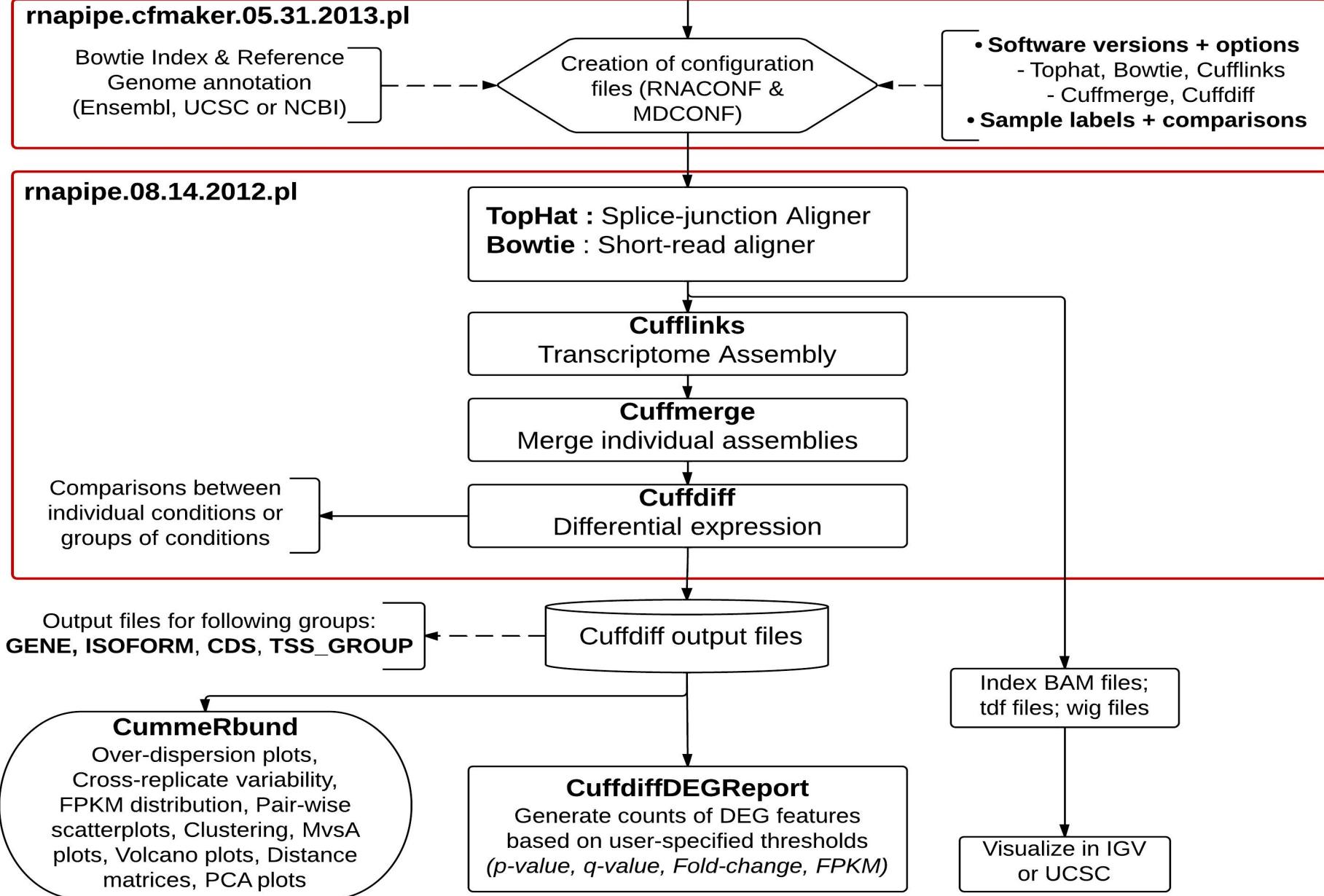
## Aggregated metrics across all samples

Alignment metrics, Insert summary metrics,  
Paired-end metrics, Strand balance,  
Coverage, Distribution across genomic  
features

# RNA-Seq analysis pipeline



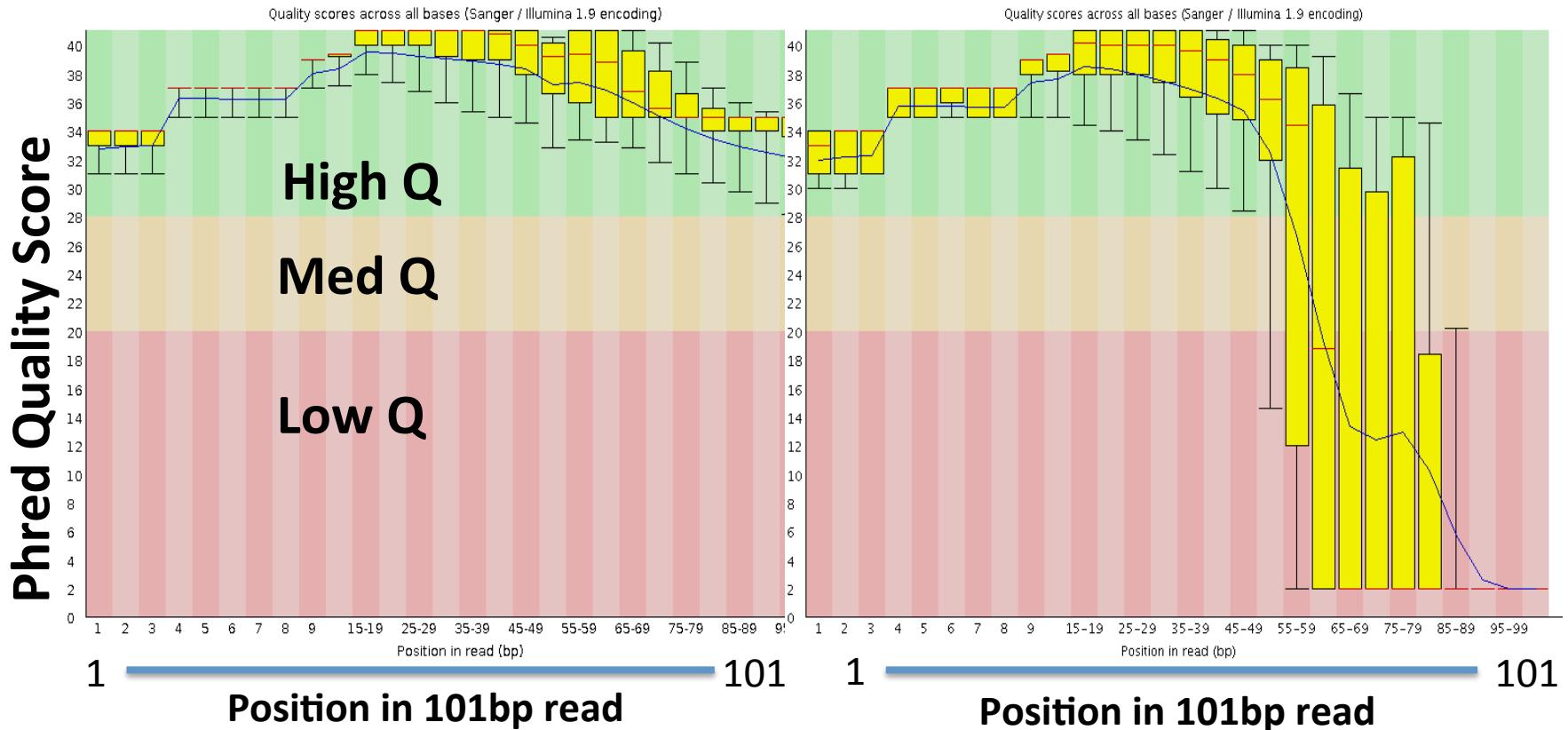
# Tuxedo RNA-Seq analysis workflow



# Outline

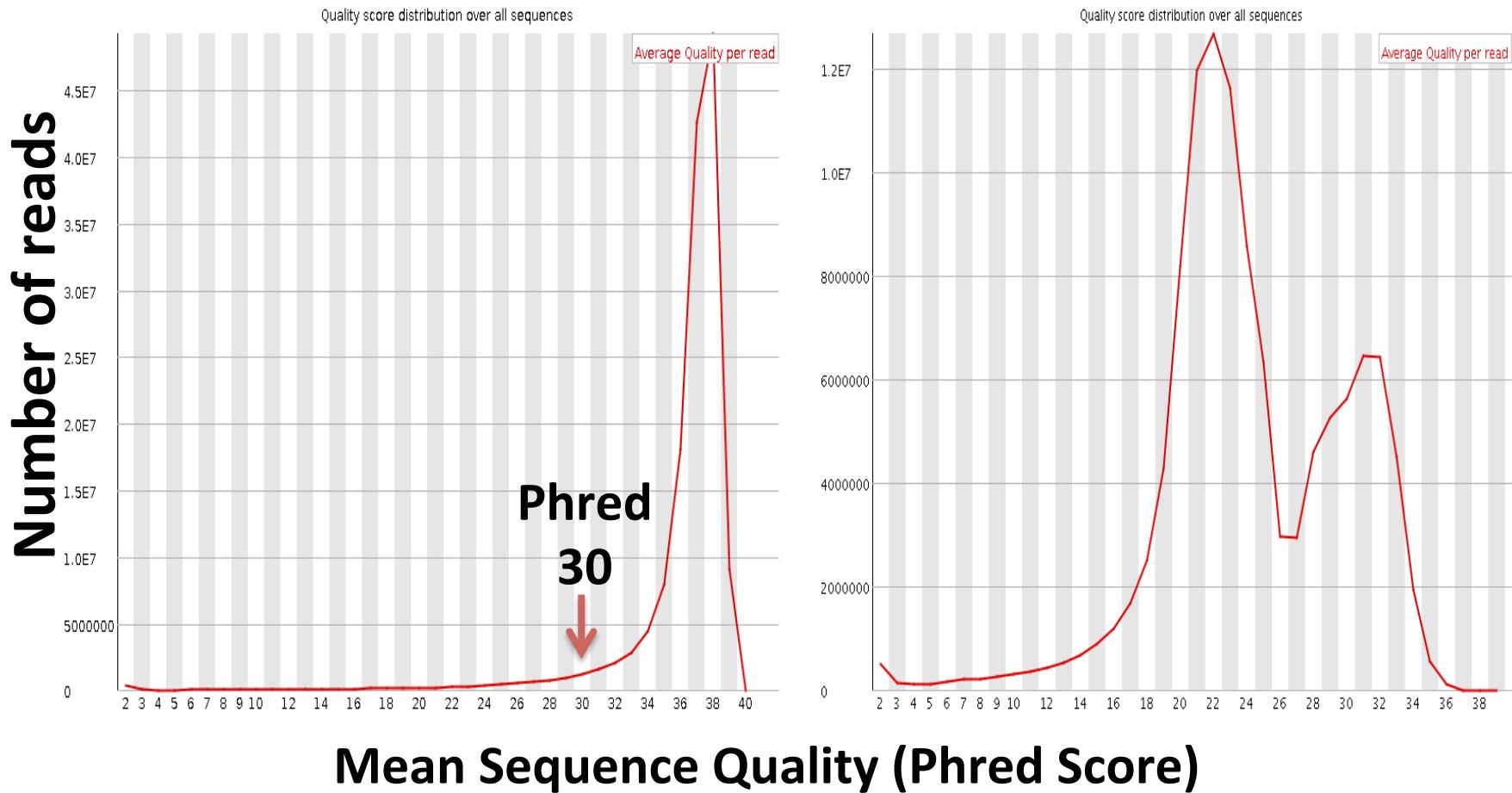
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# Per base sequence quality



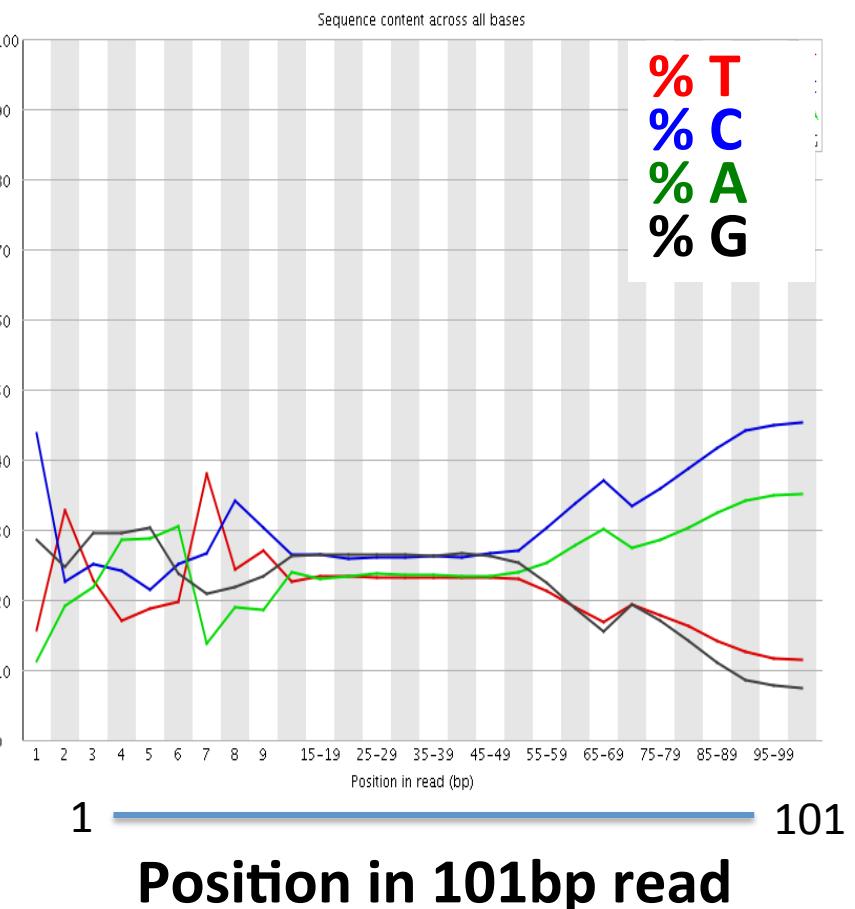
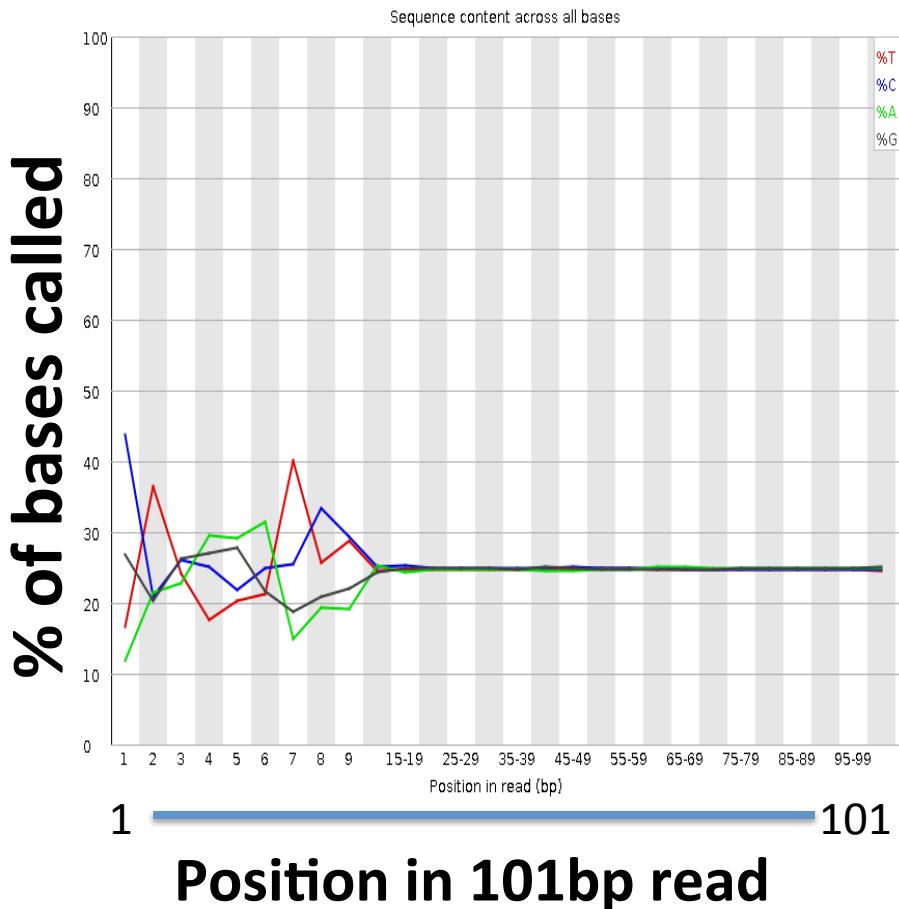
***Output from Pre-alignment QC workflow***

# Per sequence quality scores



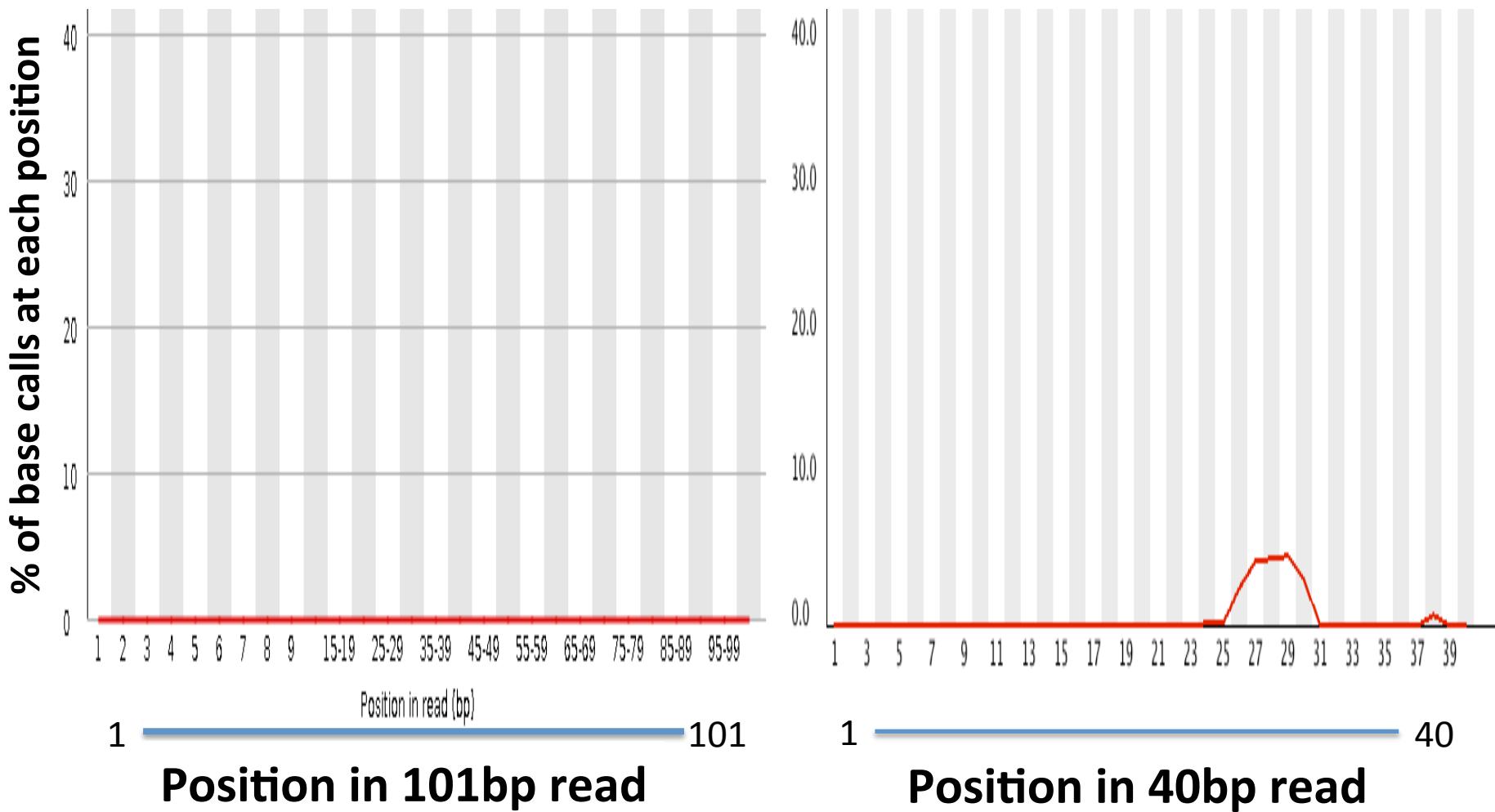
***Output from Pre-alignment QC workflow***

# Per base sequence content



***Output from Pre-alignment QC workflow***

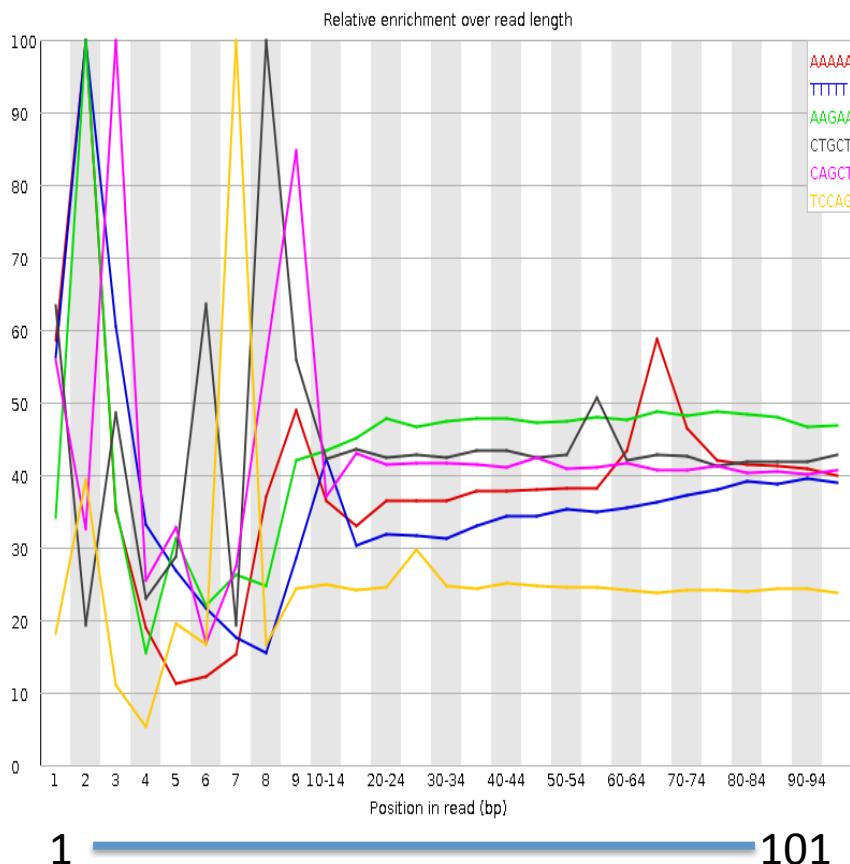
# Per base N content



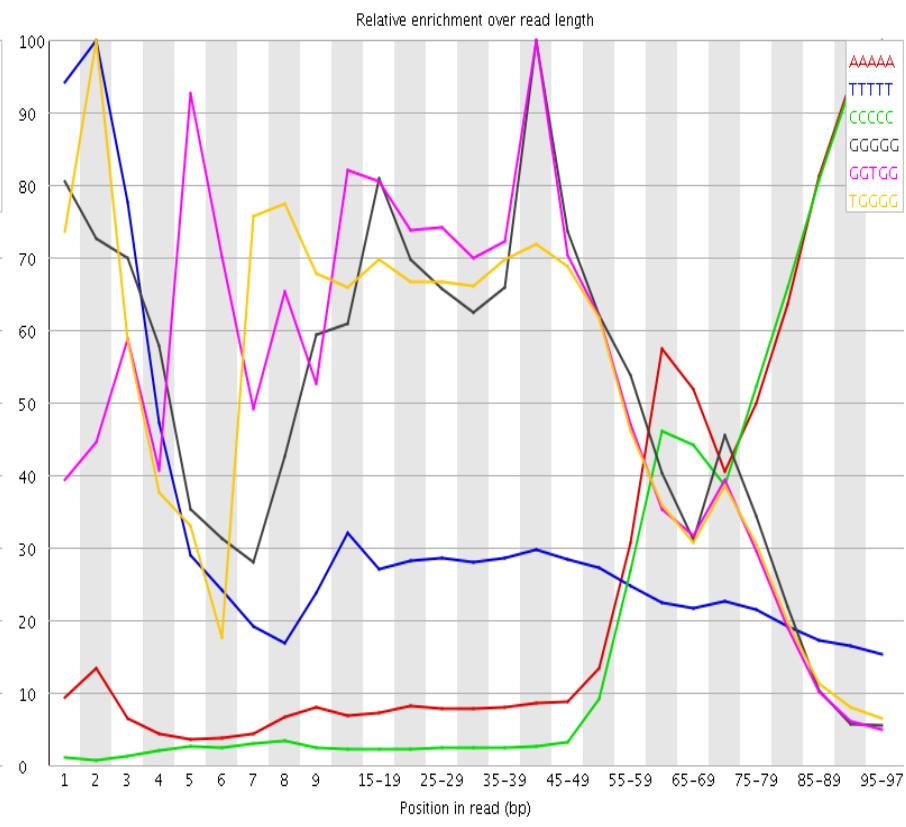
*Output from Pre-alignment QC workflow*

# Kmer content

Relative enrichment



**Position in 101bp read**



**Position in 101bp read**

***Output from Pre-alignment QC workflow***

# FastQC summary: untrimmed Fastq files

# FastQC summary: trimmed Fastq files

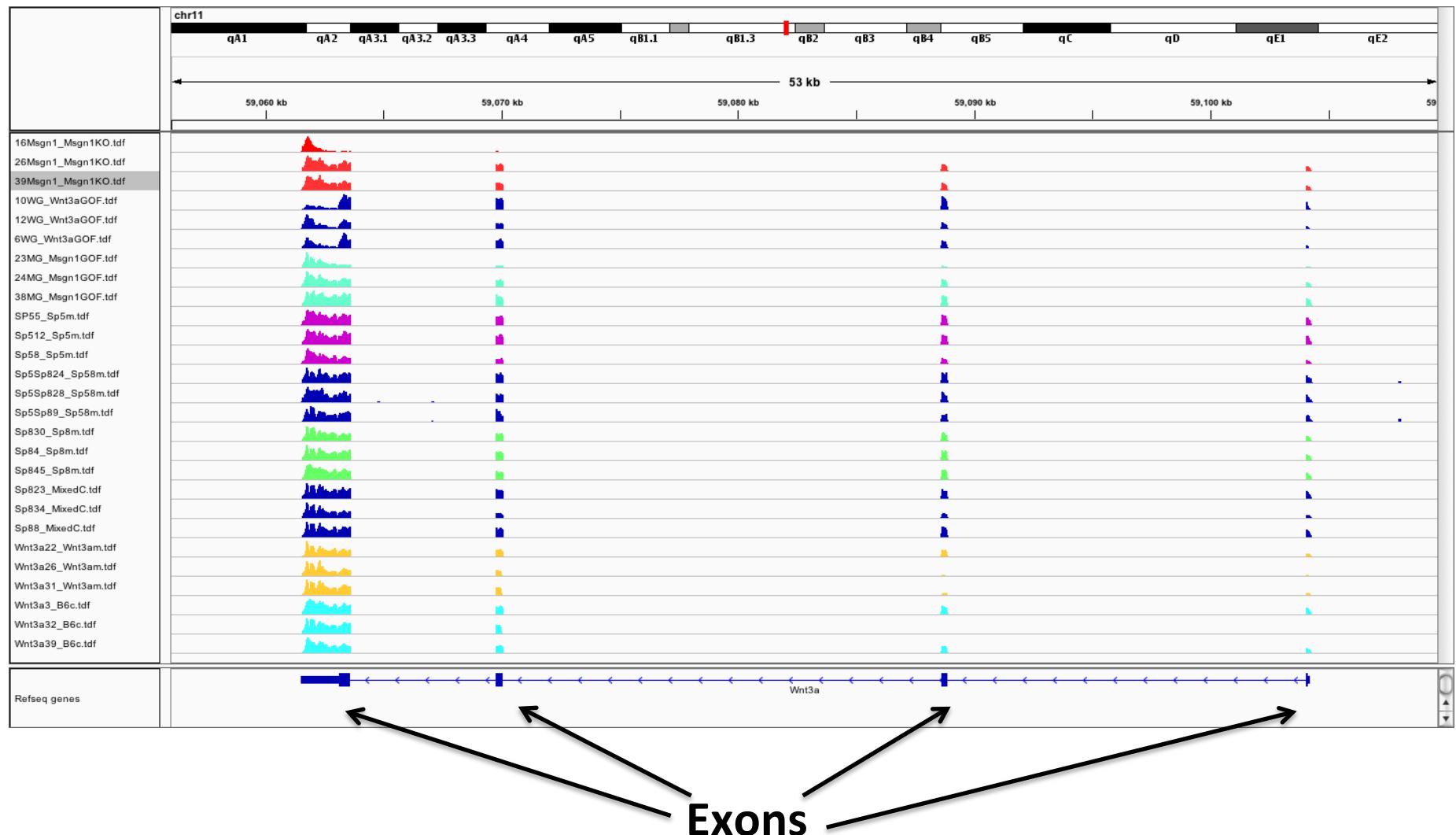
Sample	Per base sequence quality	Per sequence quality scores	Per base sequence content	Per base GC content	Per sequence GC content	Per base N content	Sequence Length Distribution	Overrepresented sequences	Kmer Content
10xWG-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	WARN	FAIL	
10xWG-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	WARN	FAIL	
12WG-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	WARN	FAIL	
12WG-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	WARN	FAIL	
16Msgn1-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	WARN	FAIL	
16Msgn1-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	WARN	FAIL	
23MG-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
24MG-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
24MG-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
26Msgn1-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
26Msgn1-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
38MG-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
38MG-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
39Msgn1-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
39Msgn1-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
3FLef1minusDox1-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	PASS	
3FLef1minusDox1-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	PASS	PASS	
3FLef1minusDox3-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	PASS	
3FLef1minusDox3-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	PASS	PASS	
3FLef1minusDox5-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
3FLef1minusDox5-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	PASS	PASS	
3FLef1plusDox1-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	PASS	
3FLef1plusDox1-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	PASS	FAIL	
3FLef1plusDox2-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	FAIL	
3FLef1plusDox2-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	PASS	FAIL	
3FLef1plusDox3-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	FAIL	
3FLef1plusDox3-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	PASS	FAIL	
6WG-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
6WG-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
FSp5minusDox2-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
FSp5minusDox2-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
FSp5minusDox3-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
FSp5minusDox3-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
FSp5minusDox4-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
FSp5minusDox4-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
FSp5plusDox1-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
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FSp5plusDox4-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
FSp5plusDox4-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
Sp12-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
Sp12-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
Sp55-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
Sp55-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
Sp58-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
Sp58-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	PASS	FAIL	
Sp5Sp824-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
Sp5Sp824-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
Sp5Sp828-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
Sp5Sp828-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
Sp5Sp889-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
Sp5Sp889-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
Sp82-Read #1	PASS	PASS	WARN	PASS	PASS	PASS	PASS	FAIL	
Sp82-Read #2	PASS	PASS	WARN	PASS	PASS	PASS	PASS	FAIL	
Sp823-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
Sp823-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
Sp830-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
Sp830-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
Sp834-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
Sp84-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
Sp84-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
Sp88-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
Sp88-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
Wnt13a2-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
Wnt13a2-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
Wnt13a6-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
Wnt13a6-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
Wnt13a9-Read #1	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
Wnt13a9-Read #2	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	
Wnt13a9-Read #3	PASS	PASS	PASS	WARN	WARN	PASS	PASS	WARN	

PASS

WARN

FAIL

# Visualization of BAM files



# Post-alignment QC: Picard metrics

## ➤ Alignment summary metrics

SAMPLE	TOTAL READS	ALIGNED READS	MISMATCH RATE	INDEL RATE	READ LENGTH	% READS ALIGNED IN PAIRS	STRAND BALANCE	% CHIMERAS
SAMPLE 1 READ 1	121,264,839	113,207,834	0.0024	0.0001	80	0.97	0.50	0.002
SAMPLE 1 READ 2	120,622,996	112,611,702	0.0019	0.0001	80	0.98	0.50	0.002
SAMPLE 1 PAIR (READ1 + READ2)	241,887,835	225,819,536	0.0022	0.0001	80	0.97	0.50	0.002

# Post-alignment QC: Picard metrics

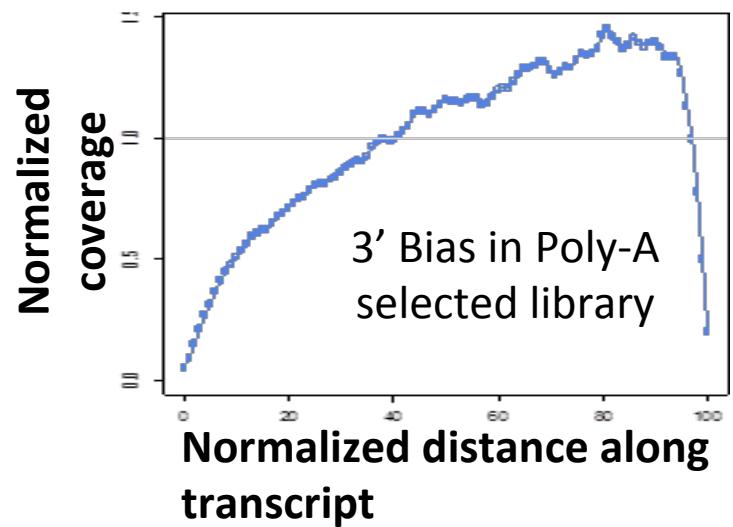
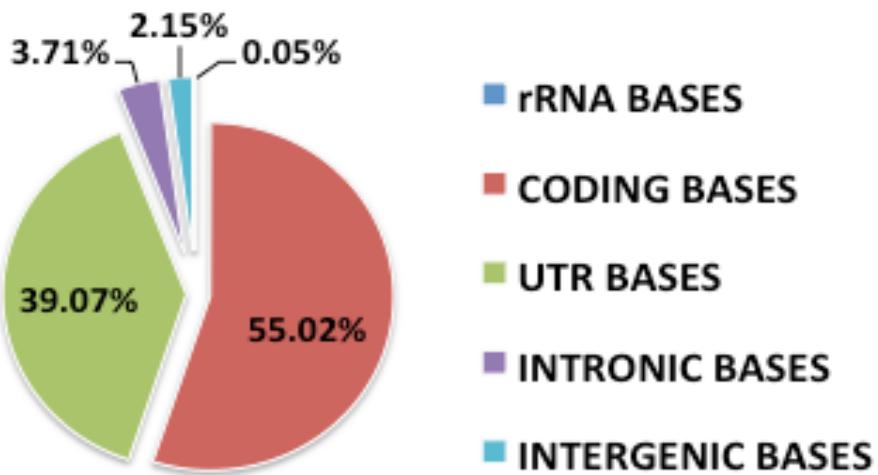
## ➤ Insert size metrics

SAMPLE	MEDIAN INSERT SIZE	MEDIAN ABSOLUTE DEVIATION	MEAN INSERT SIZE	STANDARD DEVIATION	TOTAL READ PAIRS	PAIR ORIENTATION
SAMPLE 1	189	80	229.66	189.05	116,986,895	FR
SAMPLE 2	161	55	184.31	120.28	146,691,871	FR
SAMPLE 3	159	52	181.34	112.64	131,356,931	FR

# Post-alignment QC: Picard metrics

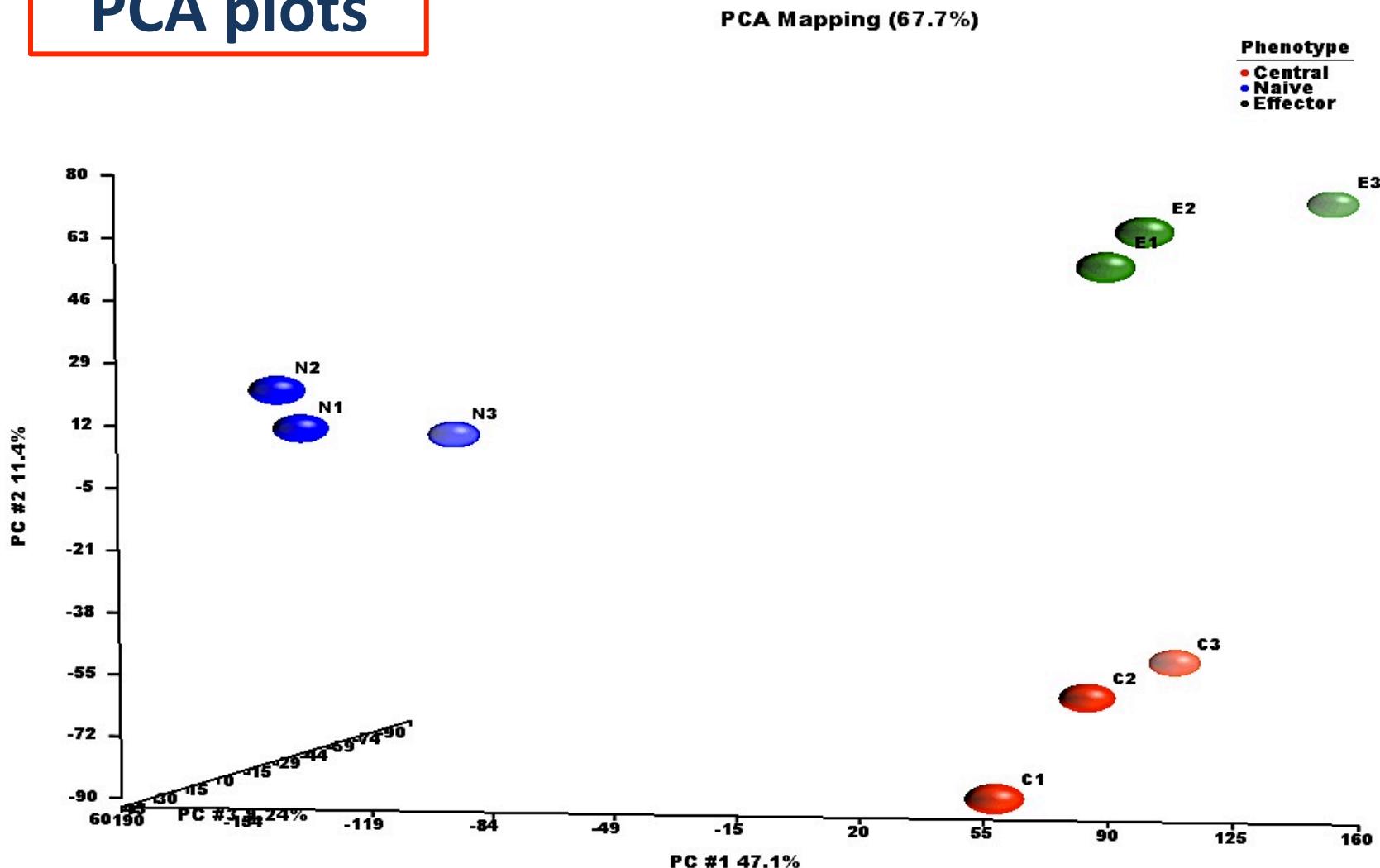
## ➤ RNASeq metrics

SAMPLE	rRNA BASES	CODING BASES	UTR BASES	INTRONIC BASES	INTERGENIC BASES	mRNA BASES	MEDIAN CV COVERAGE	5' BIAS	3' BIAS	5' TO 3' BIAS
SAMPLE 1	0.05%	55.02%	39.07%	3.71%	2.15%	94.09%	0.509	0.319	0.373	0.696
SAMPLE 2	0.11%	48.41%	47.16%	2.19%	2.14%	95.57%	0.689	0.169	0.600	0.275
SAMPLE 3	0.27%	45.47%	49.79%	2.12%	2.35%	95.26%	0.799	0.112	0.726	0.165



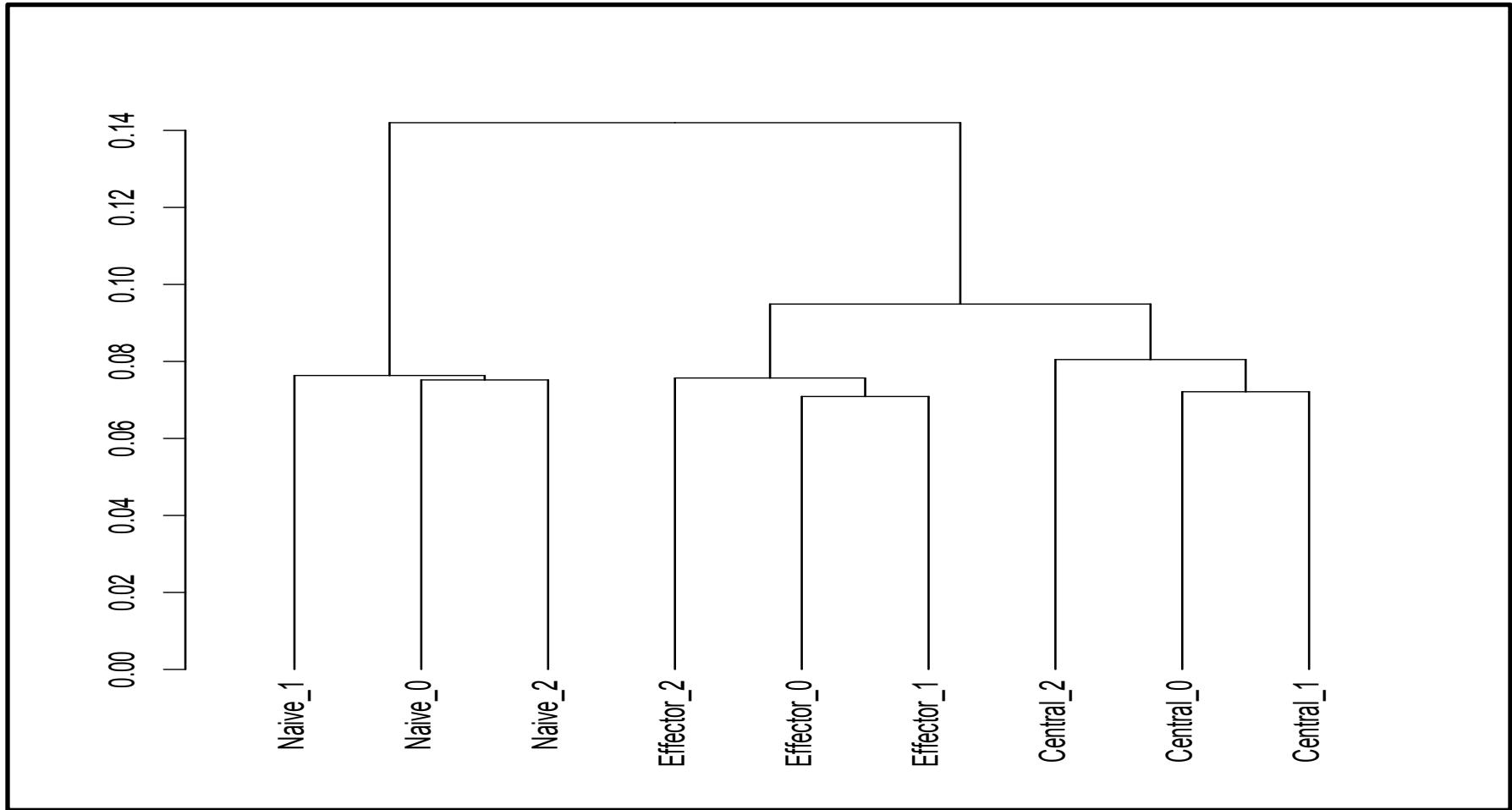
# Visuals of downstream analysis

## PCA plots



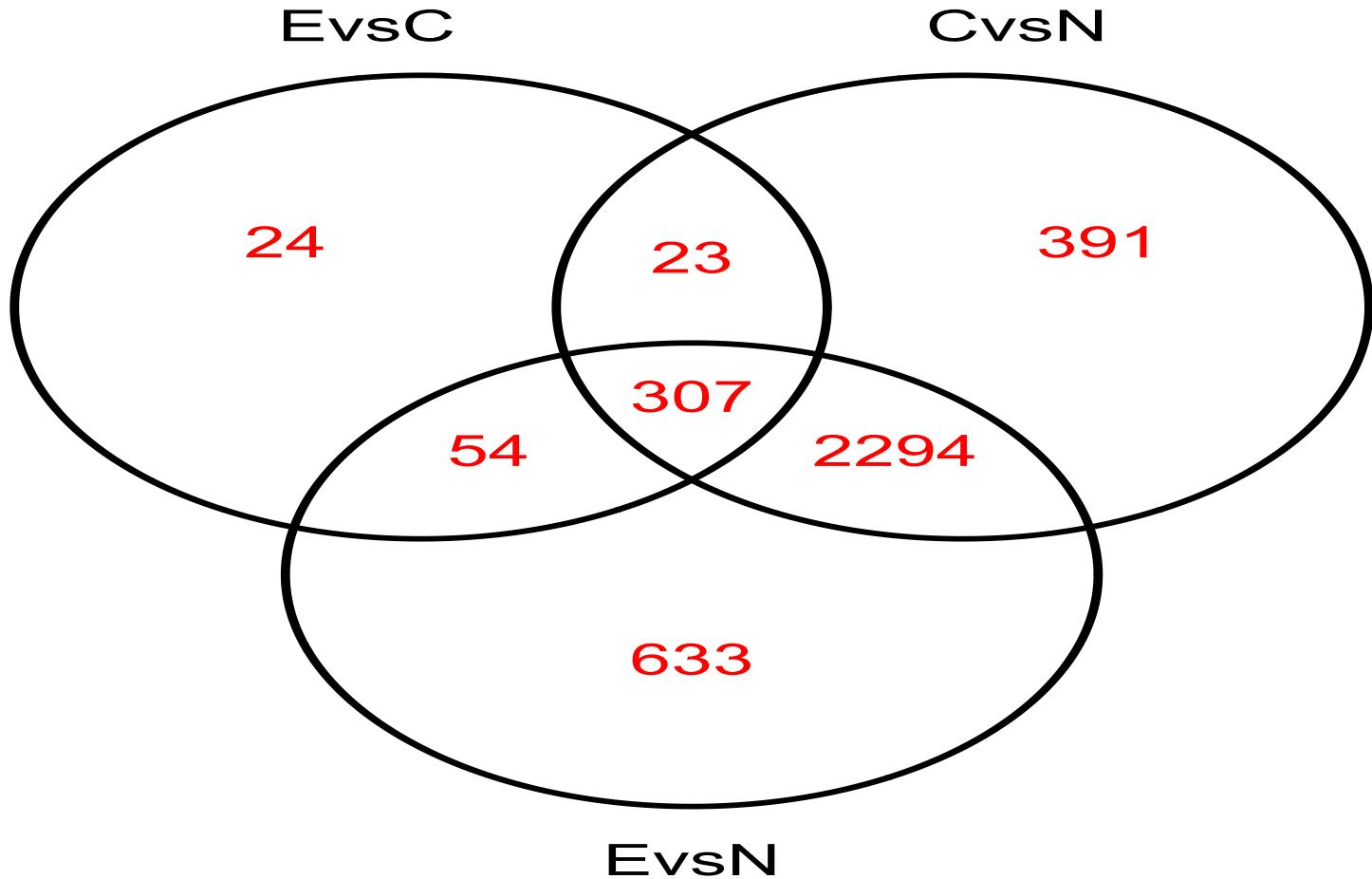
# Visuals of downstream analysis

## Dendrograms



# Visuals of downstream analysis

## Venn diagrams



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Information about Bioinformatics Presentations presented by CCRIFX

### Presentations

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Name	Summary	Downloads
RNA-Seq Workflows PPT	Powerpoint slides on RNA-Seq workflows presented by CCRIFX at the BTEP RNA-Seq workshop on 02/18/2014	 <a href="#">Presentation.pdf</a>
ChIP-Seq PPT BTEP Workshop	PowerPoint on ChIP-Seq Analysis by CCRIFX at BTEP workshop on 11/21/2013	 <a href="#">Presentation.pdf</a>

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**<http://ccrifx.cancer.gov/apps/site/presentations>**

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## Workflows for Bioinformatics Analysis

Information about Bioinformatics Workflows used by CCRIFX

### Workflows for Bioinformatics Analysis

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Name	Summary	Downloads
Tuxedo RNA-Seq analysis	A set of two scripts that automate the execution of all steps of the Tuxedo pipeline (mapping, transcriptome assembly, differential expression estimates) using configuration files	 <a href="#">Flowchart.pdf</a>  <a href="#">userDocumentation.pdf</a>
Post-alignment RNA-Seq Quality Control	Automated generation of a series of quality control metrics for RNA-Seq data using BAM files as input. Various Picard and RNA-SeQC alignment metrics are computed and user-friendly aggregated reports are compiled from generated metrics. Supports integrated and automated pre-processing of BAM files to make them suitable for QC modules to work correctly	 <a href="#">Flowchart.pdf</a>  <a href="#">userDocumentation.pdf</a>
Pre-alignment Quality Control	Workflow to merge Fastq files based on Illumina naming convention, followed by automated job submission for running FastQC on merged FastQ files. Optionally, trimming and/or adapter filtering can be carried out using Trimmomatic.	 <a href="#">Flowchart.pdf</a>  <a href="#">userDocumentation.pdf</a>

[ccrifx.cancer.gov/apps/site/workflows\\_for\\_bioinformatics\\_analysis](http://ccrifx.cancer.gov/apps/site/workflows_for_bioinformatics_analysis)

# Visit our website for more information



The screenshot shows the homepage of the CCR IFX INFORMATICS website. At the top right is a 'Login →' link. Below it is a navigation bar with links: Home, Resources, FAQ, Staff, Projects, Examples, Insights (which is highlighted in red), and Services. The main header features the CCR IFX logo and the text 'Analysis Support Process' with the subtitle 'Supporting your analysis needs'. The main content area has a section titled 'Analysis Support Process' with a detailed description of the core's role and support process. It also lists four basic stages: Definition Phase, Prioritization Phase, and Execution Phase. To the right is a 'Site Search' bar with a magnifying glass icon and a 'Useful Information' sidebar containing links to various support topics.

**Analysis Support Process**

The CCRIFX bioinformatics core is a shared resource serving several hundred scientists and investigators working for the NCI Center for Cancer Research. Even so, we treat every investigator as a long-term collaborator with individual needs and deadlines. As a shared resource, the core must also be able to work well and fairly with many investigators and employ a support process to make this happen.

The support process follows four basic stages :

**Definition Phase** – a support project request is submitted through the website ([Request Project](#)) and kicks-off the analysis support. During this phase, analysts meet with investigators and scientists to clarify the nature of the request and address open questions that would slow or preclude a timely response to the request. Some projects will be very straight forward and move ahead quickly, while other requests may involve a significant amount of new exploration and require more time to define.

**Prioritization Phase** – As a shared resource, CCR has established a request prioritization committee (CCRIFX-RPC) that helps to prioritize requests for efficient and best response. The RPC committee is kept aware of new requests and completed definitions in order to keep the process moving ahead in a timely fashion. Dr. David Goldstein heads the CCRIFX-RPC and investigators are encouraged to contact him regarding questions about the prioritization process.

**Execution Phase** – Once approved and prioritized, the request moves ahead for subsequent analysis by the core

**Site Search**

**Useful Information**

- > Analysis Support Process
- > Office Hours and Support
- > Support Request Prioritization
- > Education and Training
- > Workflows for Bioinformatics Analysis
- > Scientific Discussions

[ccrifx.cancer.gov/apps/site/analysis\\_support\\_process](http://ccrifx.cancer.gov/apps/site/analysis_support_process)

# List of RNA-Seq Bioinformatics tools

Tool category	Tools
QC, filtering, pre-processing	<b>FastQC</b> , <b>Trimmomatic</b> , <b>RNA-SeQC</b> , <b>Picard</b> , Cutadapt, FASTX, Flexbar, htSeqTools, PRINSEQ, qrqc, RSeQC, SAMstat, SEECER, ShortRead
Alignment	<b>Unspliced</b> : BFAST, <b>Bowtie</b> , BWA, SOAP, Maq, Mosaik, NovoAlign, SHRIIMP, Stampy <b>Spliced (known)</b> : <b>TopHat</b> , Erange, RUM, RNASEQR, SpliceSeq <b>Spliced (de novo)</b> : <b>TopHat</b> , GSNAp, HMMSplicer, MapSplice, QPALMA, SpliceMap
Quantitative analysis & differential expression	<b>Cufflinks</b> , <b>DESeq</b> , <b>EdgeR</b> , DEGSeq, DEXSeq, DiffSplice, BaySeq, Alexa-Seq, BitSeq,, ERANGE, eXpress, RSEM, SpliceTrap
Workbench (integrated pipeline solutions)	<b>Commercial</b> : <b>Partek</b> , <b>Genomics</b> , Avadis NGS, <b>CLC</b> , DNASTAR, GeneSpring GX <b>Open Source</b> : <b>Galaxy</b> , Taverna, GenePattern, ArrayExpressHTS, easyRNASeq, MeV
Alternative splicing analysis	MISO, <b>Cufflinks/Cuffdiff</b> , DEXseq, SAJR
Fusion genes/chimeras/translocation finders/structural variations	BreakDancer, ChimeraScan, <b>FusionMap</b> , FusionSeq, SOAPFuse, SOAPfusion, <b>Tophat-Fusion</b> , <b>DeFuse</b>
Visualization tools	<b>CummeRbund</b> , <b>IGV</b> , IGB, MapView, Tablet, Savant, <b>SeqMonk</b> , Gbrowse, EagleView
Transcriptome assembly	<b>Genome-guided</b> : Cufflinks, iReckon, IsoInfer, RNAeXpress, Scripture <b>Genome-independent</b> : Oases, Rannotator, SOAPdenovo, Trans-ABYSS, Trinity, Velvet

[http://en.wikipedia.org/wiki/List\\_of\\_RNA-Seq\\_bioinformatics\\_tools](http://en.wikipedia.org/wiki/List_of_RNA-Seq_bioinformatics_tools)

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  - GenomeQuest: [http://wiki.genomequest.com/index.php/RNA\\_Seq](http://wiki.genomequest.com/index.php/RNA_Seq)
- **Blogs**
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*Thank You !*

# Extra slides

# RNA-Seq: Advantages over microarrays

- Higher resolution, more sensitivity
- High signal-to-noise ratio
- Potentially unlimited dynamic range of expression with absolute rather than relative values.
- Requires less RNA sample & results highly reproducible for both technical & biological replicates
- No a-priori knowledge of genome content required
- Investigation of both known and novel transcripts
- Alternative splicing patterns, RNA editing events

# Applications of RNA-Seq platform

- Relative expression analyses
- Characterization of alternative splicing patterns
- Discovery of novel transcripts and isoforms
- Small RNA profiling
- Allele-specific & strand-specific expression
- Exploration of non-model-organism transcriptomes
- Mapping transcription start sites
- Gene fusion detection
- Profiling low-quantity RNA samples
- RNA editing